



Published in final edited form as:

*Nutr Clin Care*. 2004 ; 7(2): 56–68.

## Probiotics and Medical Nutrition Therapy

**Amy C. Brown, Ph.D., R.D. and Ana Valiere, M.S.**

*Dr. Brown and Ms. Valiere are with the Department of Human Nutrition, Food & Animal Sciences, University of Hawaii at Manoa, 1955 East West Road, Room 216, Honolulu, HI 96822 USA.*

### Abstract

Probiotics have been defined by The Food Agricultural Organization/World Health Organization (FAO/WHO) as “live microorganisms which when administered in adequate amounts confer a health benefit to the host.” They have been used for centuries in the form of dairy-based fermented products, but the potential use of probiotics as a form of medical nutrition therapy has not received formal recognition. A detailed literature review (from 1950 through February 2004) of English-language articles was undertaken to find articles showing a relationship between probiotic use and medical conditions. Medical conditions that have been reportedly treated or have the potential to be treated with probiotics include diarrhea, gastroenteritis, irritable bowel syndrome, and inflammatory bowel disease (Crohn’s disease and ulcerative colitis), cancer, depressed immune function, inadequate lactase digestion, infant allergies, failure-to-thrive, hyperlipidemia, hepatic diseases, *Helicobacter pylori* infections, genitourinary tract infections, and others. The use of probiotics should be further investigated for possible benefits and side-effects in patients affected by these medical conditions.

### Keywords

probiotic; medical nutrition therapy; diet; disease

## PROBIOTICS AND HEALTH

Probiotics are nonpathogenic organisms (yeast or bacteria, especially lactic acid bacteria) in foods that can exert a positive influence on the host’s health.<sup>1</sup> The theory is that live microorganisms within food or in the form of a supplement improve the microbial balance of the intestinal tract.<sup>2</sup> The most commonly consumed probiotics are fermented dairy products such as yogurt and buttermilk. Probiotic therapy is not a new idea; it dates back almost 100 years to Elie Metchnikoff, who suggested that Bulgarian peasants lived longer lives because of their yogurt consumption. In the 1930s, a Japanese physician, Minoru Shirota, suggested that the right mix of bacteria in the gut could prevent disease. Miso soup, made from fermented soybean paste, is a staple of the Okinawan diet.

It has been suggested that disrupting the delicate balance in the gastrointestinal tract can contribute to diarrhea (antibiotic-associated diarrhea, traveler’s diarrhea, intestinal infections),<sup>1,3</sup> gastroenteritis, constipation, irritable bowel syndrome, inflammatory bowel disease (Crohn’s disease and ulcerative colitis), food allergies, and certain cancers.<sup>4–6</sup> On the contrary, a balanced or “normal” enteric flora may competitively exclude possible pathogenic organisms, stimulate the intestinal immune system,<sup>7,8</sup> and produce nutrients and other substances such as short-chain fatty acids, vitamins,<sup>9</sup> amino acids (arginine, cysteine, and glutamine), polyamines, growth factors, and antioxidants.<sup>7</sup>

## PROBIOTIC SUPPLEMENTATION STUDIES

Studies show that commercial probiotic consumption often increases specific intestinal microflora, but usually not the total count of bacteria found in the intestine. It is also evident

in the majority of reported research cases that specific bacteria do not increase unless subjects consume very high dosages of probiotics in the form of supplements, not those naturally found in foods.

One of the early double-blind, placebo-controlled studies (20 men; 10/group) utilizing a commercially available probiotic was conducted by Spanhaak and associates,<sup>10</sup> who reported an increase in *Lactobacillus* count. Benno and Mitsuoka<sup>11</sup> reported that *Bifidobacterium longum* administered as a pharmaceutical in adults resulted in higher fecal bifidobacterial and lower clostridial counts, lower fecal pH, and lower fecal ammonia concentrations. A study of 64 adults by Ling et al.<sup>12</sup> showed that consumption of *Lactobacillus GG* resulted in higher fecal counts of *Lactobacillus GG*, decreased fecal beta-glucuronidase, nitroreductase, and glycocholic acid hydrolase activities. There was also a decrease in urinary p-cresol excretion, which is a product of colonic *Bacteriodes fragilis*. Studies in which bottle-fed infants were given an inoculum of *Bifidobacterium bifidum* and compared with control infants showed that *B. bifidum* did appear in the stools of infants in the treatment groups.<sup>13,14</sup>

## SURVIVAL OF GASTROINTESTINAL MICROFLORA

The benefit of probiotics is based on the survival of these bacterial or yeast cultures in the gastrointestinal tract and the resulting effects they might generate to overall health. Studies evaluating how well probiotics fare through the gastrointestinal tract show that about 10–30% of probiotics survive. This depends on a number of variables, including the type of probiotic.<sup>15,16</sup>

Probiotic bacteria's survival in food products also varies according to strain differences, the product's storage conditions, the chemical composition of the product to which the probiotics are added, and other interactions between the components of the product.<sup>17</sup> Table 1<sup>3,18</sup> lists the most commonly used bacterial strains for probiotic purposes. Certain strains of lactic acid bacteria, such as *Lactobacillus plantarum* and *Lactobacillus rhamnosus*, have proved to be more effective in preserving the microbial balance of the colon, preserving the key nutrients produced by the bacteria, eliminating toxic components from food, protecting from decay, and obliterating pathogens.<sup>7</sup> In addition to the influence of various strains, several other characteristics essential for probiotics are adherence to cells, gastric acid and bile stability, production of antimicrobial substances, and activity against pathogenic bacteria.<sup>19</sup> Probiotics, regardless of their strain, must be resistant to acid pH and biliary acids in order to colonize the intestine.<sup>3</sup>

## PROBIOTIC USE IN MEDICAL CONDITIONS

Even if the probiotics are viable, certain medical conditions and their relationships to probiotics are highly controversial. Their possible impact on gastrointestinal health is just beginning to be understood, and it must be remembered that multiple variables exist, along with the possibility of remission due to unknown factors. What follows is a brief review of the possible beneficial effect of probiotics on diarrhea, gastroenteritis, irritable bowel syndrome, inflammatory bowel disease (Crohn's disease and ulcerative colitis), cancer, the immune system, lactose digestion, infant allergies, failure-to-thrive, hyperlipidemia, hepatic disease, *Helicobacter pylori* infections, genitourinary tract infections, and other medical conditions (Table 2).<sup>20</sup>

## DIARRHEA

Probiotics have been reported to treat three types of diarrhea: antibiotic-associated, traveler's, and infectious. The most common side effect of antibiotic therapy is diarrhea, which occurs in about 20% of patients due to a disruption in the balance of the endogenous flora in the colon.

<sup>1,3</sup> Antibiotic use can encourage the growth of pathogenic bacteria, specifically *Clostridium difficile* and *Klebsiella oxytoca*.<sup>1,3</sup> There have been several clinical trials attempting to determine the efficacy of administering probiotics to patients experiencing antibiotic-associated diarrhea (Table 3).<sup>21</sup> Three randomized, double-blind, controlled studies reduced the rate of occurrence of antibiotic-associated diarrhea using orally administered *Saccharomyces boulardii*. In animal models, *S. boulardii* has also been shown to decrease *C. difficile*.<sup>3</sup> *Lactobacillus rhamnosus* GG, another probiotic strain showing promise in treating AAD,<sup>3,21</sup> has been shown to colonize the colon of patients receiving erythromycin, penicillin, and ampicillin therapy.<sup>3</sup> The effects of other probiotic strains on antibiotic-associated diarrhea have been mixed. For example, Gotz et al. reported that a mixture of *Lactobacillus acidophilus* significantly prevented diarrhea, but two other studies show that the same mixture did not significantly prevent diarrhea (Table 3). Marteau et al. suggested that this could be due to differences in probiotic preparation and in Table 3<sup>1</sup> he shows a summary of different probiotics.<sup>1,21</sup>

Traveler's diarrhea also occurs in about half of the people who travel to underdeveloped or high-risk countries, and can range from mild to severe.<sup>1</sup> At least eight clinical trials have been conducted testing the use of orally administered probiotics to prevent traveler's diarrhea (Table 4).<sup>1</sup> A randomized study by Black and colleagues used a mixture of probiotic strains (*L. acidophilus*, *L. bulgaricus*, *B. bifidum*, and *S. thermophilus*) or a placebo to treat 94 Danish tourists traveling to Egypt for 2 weeks.<sup>1,22,23</sup> The incidence of traveler's diarrhea in the tourists was reduced significantly from 71% (placebo group) to 43% (probiotic group).<sup>1,22,23</sup> The oral administration of *L. rhamnosus* GG also resulted in a decreased occurrence of traveler's diarrhea, with the most significant study using 245 New Yorkers that traveled to a developing nation (3.9% probiotic group versus 7.4% placebo group).<sup>1,24</sup> Many other probiotic strains have had insignificant results (Table 4).<sup>1</sup>

The range of causes for intestinal infections causing diarrhea vary greatly. Some of the possible pathogens include *Shigella*, *Salmonella*, *Campylobacter*, *Colostridium difficile*, rotavirus, enterotoxigenic *Escherichia coli*, and *Helicobacter pylori*.<sup>1,25</sup> Infection with rotavirus is the most common cause of severe diarrhea in children, and probiotic therapy (*Lactobacillus GG*) has been shown to be effective in reducing the duration of rotavirus enteritis infection.<sup>26</sup> A meta-analysis suggested that *Lactobacillus* is an effective treatment for children with acute infectious diarrhea.<sup>27</sup>

No significant benefit was seen regarding the use of *L. acidophilus* and *L. bulgaricus* for people with *E. coli* infection in a double-blind, placebo-controlled study.<sup>28</sup> Also, no benefit was seen with the use of *E. faecium* strain SF68 for adults with *Vibrio cholerae* and *E. coli* infection.<sup>29</sup> Upon colonization of the gastric mucosa, *H. pylori* infection can lead to gastritis, duodenal and gastric ulcers, and even some malignancies.<sup>1,25</sup> Some *Lactobacillus* strains have shown antagonistic actions against *H. pylori* in vitro<sup>56,30</sup> and in a gnotobiotic murine model.<sup>31,32</sup> The few human studies regarding *Lactobacillus* and *H. pylori* infection have had conflicting results.<sup>25</sup> Guandalini et al.<sup>26</sup> found that *Lactobacillus* GG helped their child subjects recover more rapidly from diarrhea caused by rotavirus enteritis; however, they also stated that information is limited as to the role of this probiotic in non-rotaviral diarrheal episodes.

## GASTROENTERITIS

Inflammation of the mucous membrane of the intestines, or gastroenteritis, is the main cause of acute diarrhea, which generally lasts a few days.<sup>1</sup> The inflammation can result from viral pathogens, bacterial pathogens, or parasites; in children, however, it usually results from rotavirus infection. Oral rehydration therapy is the most common therapy, but does not reduce the duration of diarrhea.<sup>1,3,25</sup> Several controlled, randomized trials have shown a shortened

duration of rotavirus-associated diarrhea and gastroenteritis in infants with the use of probiotics, especially with *L. rhamnosus* GG (Table 5).<sup>1,25</sup> In addition, several nonrandomized trials have indicated a preventive effect of some fermented products on the incidence of diarrhea in children.<sup>33,34</sup> The results of adult gastroenteritis studies with probiotics have been less significant.<sup>1</sup>

## IRRITABLE BOWEL SYNDROME

Symptoms most often associated with irritable bowel syndrome (IBS) may include, but are not limited to constipation and/or diarrhea, abdominal pain, flatulence, and bloating. In a randomized, controlled clinical trial conducted on 60 patients, Nobaek et al.<sup>35</sup> reported that orally administered *L. plantarum* reduced pain and flatulence. In a small study of IBS sufferers given *L. plantarum*, participants demonstrated a small, yet significant decrease in abdominal pain.<sup>36</sup> Kim et al.<sup>37</sup> administered a probiotic formulation (VSL#3 twice daily for 8 weeks) to 25 patients with IBS in a randomized, controlled trial. There was a significant reduction in abdominal bloating, indicating that VSL#3 may assist patients with IBS.<sup>37</sup> In another study conducted by Brigidi et al.<sup>38</sup> 10 patients with IBS were given VSL#3, resulting in fecal microbiota showing increased concentrations of *Lactobacilli*, *Bifidobacteria*, *Streptococcus thermophilus*, and fecal beta-galactosidase.<sup>38</sup>

## INFLAMMATORY BOWEL DISEASE

Inflammatory bowel disease (IBD) encompasses intestinal inflammatory diseases such as Crohn's disease and ulcerative colitis. These diseases all result from unknown origins, but intestinal flora disturbances<sup>39,40</sup> and a defective mucosal barrier<sup>41</sup> have been hypothesized as contributing factors. Intestinal infections sometimes mimic IBD symptoms but disappear with antibiotic therapy. There have been several IBD animal studies using probiotics with promising results<sup>1,39</sup>; however, only a few trials have been conducted in humans.<sup>23,42</sup> A study involving 14 children with Crohn's disease receiving *L. rhamnosus* GG found an increase in immunoglobulin A immune response.<sup>43</sup> This study has been criticized for its small sample size and short duration of 10 days. Additionally, *S. boulardii* has been shown to significantly reduce disease activity and the frequency of bowel movements in 20 Crohn's disease patients.<sup>44</sup> *S. boulardii* was also noted to reduce relapse rates and to extend remission time in patients with Crohn's disease.<sup>45</sup> Malchow<sup>46</sup> conducted a randomized, controlled clinical trial that involved 28 patients with Crohn's disease (only in the colon) who received either their standard treatment (glucocorticoids) plus a capsule containing a viable nonpathogenic *E. coli* strain Nissle 1917 or the steroid and a placebo. He found that administration of nonpathogenic *E. coli* reduced the risk for relapse and decreased the need for glucocorticoids.<sup>46</sup>

Further research is needed on probiotic treatments for ulcerative colitis.<sup>23,25</sup> A study by Venturi et al.<sup>47</sup> found that a combined probiotic preparation known as VSL#3 effectively maintained remission in mesalazine-intolerant ulcerative colitis patients. The VSL#3 preparation included four strains of lactobacilli, three strains of bifidobacteria, and *Streptococcus salivarius*. Gionchetti et al.<sup>23</sup> also used the VSL#3 combination in a study and found that it prevented the development of chronic pouchitis following pouch-anal anastomosis surgery treatment for ulcerative colitis.

## CANCER

Probiotics have been found by several researchers to decrease fecal concentrations of enzymes and secondary bile salts, and reduce absorption of harmful mutagens that may contribute to colon carcinogenesis.<sup>48</sup> Other studies suggest that normal intestinal flora can influence carcinogenesis by producing enzymes (glycosidase, B-glucuronidase, azoreductase, and nitroreductase) that transform precarcinogens into active carcinogens.<sup>12,49–52</sup> Certain

probiotics may protect the host from this activity. *L. acidophilus* and *L. casei* supplementation in humans helped to decrease levels of these enzymes, as shown by fecal specimens.<sup>53–55</sup> In animal studies, the bacterial enzymes aforementioned have been suppressed with the administration of *Lactobacillus GG*.<sup>45</sup> Other lactic acid bacteria have shown similar results; however, the relationship between enzyme activity and cancer risk needs further investigation.

Certain studies have shown an effect of probiotics on tumor growth. Burns and Rowland<sup>56</sup> suggested that increasing the amount of lactic acid bacteria in the colon decreases the ability of microflora to produce carcinogens. A randomized, controlled study by Aso and Akazan<sup>57</sup> of 48 Japanese patients demonstrated that the recurrence of bladder tumors was delayed with daily intake of *L. casei*. They performed another study that was larger (125 patients) and placebo-controlled, and found that *L. casei* reduced the recurrence of tumors in all patients except those with more than one recurrent tumor.<sup>58</sup> The hypothesis is that lactobacilli might bind mutagenic compounds in the intestine, which would reduce the absorption of these harmful mutagens.<sup>59</sup> A short-term study has confirmed this hypothesis by measuring the excretion of mutagens in urine after consumption of hamburger meals supplemented with *L. acidophilus*.<sup>60</sup> In an animal model with dimethylhydrazine (DMH)-induced colon cancer, it was shown that *Lactobacillus GG* significantly reduced the incidence of colon tumors.<sup>61</sup> Lactic acid bacteria administered to animals have been shown to prevent carcinogen-induced preneoplastic tumors and lesions.<sup>62</sup> Hirayama et al.<sup>63</sup> found that lactic acid bacteria reduced the growth and viability of the HT-29 human colon cancer line. A review article by Vanderhoof<sup>64</sup> describes a study using two carcinogens to test the effects of probiotics on the prevention of DNA damage in an animal model. The carcinogens used were N-methyl-N-nitro-N-nitrosoguanidine (MNNG) and DMH. Several probiotic strains were tested in this study, including *L. gasseri*, *L. confusus*, *S. thermophilus*, *B. breve*, *B. longum*, and *L. acidophilus*. All of the strains showed an antigenotoxic effect after MNNG administration.<sup>64</sup> Similar studies have shown that pretreatment with *L. acidophilus*, *L. confusus*, *L. gasseri*, *B. longum*, and *B. breve* inhibited DNA damage from DMH, but that only one of four *S. thermophilus* strains and only one of three *L. delbrueckii* strains were protective.<sup>48</sup> Another study found that all five of the lactic acid bacteria strains that they tested inhibited the growth of the MCF7 breast cancer cell line, although *B. infantis* and *L. acidophilus* were the most effective.<sup>65</sup>

Several mechanisms have been proposed as to how lactic acid bacteria may inhibit colon cancer; these include: enhancing the host's immune response, altering the metabolic activity of the intestinal microflora, binding and degrading carcinogens, producing antimutagenic compounds, and altering the physicochemical conditions in the colon.<sup>63</sup> Studies have suggested that the host's immune response may be stimulated by *B. infantis*, leading to tumor suppression or regression.<sup>63</sup> The metabolic activity of the intestinal microflora may also be altered with administration of lactic acid bacteria. Goldin and Gorbach<sup>66</sup> studied the effect of *L. acidophilus* on three bacterial enzymes ( $\beta$ -glucuronidase, nitroreductase, and azoreductase) in 21 volunteers for 10 days. *L. acidophilus* reduced the activity of the carcinogen-releasing bacterial enzymes. Binding and degrading carcinogens may be possible by lactic acid bacteria supplementation. The production of antimutagenic compounds in the colon has been demonstrated with *B. longum* administration in rats. Azomethane-induced colon tumor development was also suppressed with a decrease in colonic mucosal cell proliferation and tumor ornithine decarboxylase and ras-p21 activities.<sup>63</sup> The alteration of the physicochemical conditions in the colon may influence colon cancer, and Modler et al.<sup>67</sup> suggest that reducing the intestinal pH may prevent the growth of putrefactive bacteria. In a 3-month study, *L. acidophilus* and *B. bifidum* were administered to patients with colonic adenomas. The result was a decrease in fecal pH and cell proliferative activity in the upper colon.<sup>68</sup> The mechanisms of the links of probiotics to antitumor activity are not completely clear, but offer useful potential material for future cancer studies.

## IMPROVED IMMUNE SYSTEM

Probiotics have also been shown to influence several aspects of immune function. In animal and human studies with different bacteria treatments (*L. casei*, *L. acidophilus*, or *B. bifidus*), an enhanced secretory immunoglobulin (Ig) A production was observed.<sup>70</sup> *L. casei* is most effective in stimulating secretory Ig A<sup>70</sup> and increasing the systemic immune response in malnourished animals.<sup>71</sup> Another study showed that mice fed lactic acid bacteria had higher splenocyte proliferation in response to mitogens for T and B cells.<sup>69,72</sup> Several studies have shown that probiotics (*L. casei*, *L. rhamnosus* GG, and other strains) can affect cytokine production.<sup>69</sup> In addition, several studies have shown that probiotics promote a nonspecific immune response by enhancing phagocytosis of pathogens.<sup>69</sup> The mechanisms by which probiotics affect the immune system and its responses are still yet to be determined.

## IMPROVED LACTOSE DIGESTION

It is well established that lactose is digested better in fermented dairy foods such as yogurt than in non-fermented dairy products.<sup>73-78</sup> Despite being studied extensively, the mechanisms involved have not clearly been deciphered. The viability of lactic acid bacteria has been proposed to be involved with this improved digestibility because pasteurization is known to reduce lactose digestibility.<sup>1,74,76,78</sup> In 1991, Martini and colleagues<sup>79</sup> proposed that the improved digestibility was partially due to the activity of bacterial enzymes ( $\beta$ -galactosidases) produced from the two lactic acid bacteriums used to ferment milk to yogurt (*Streptococcus salivarius* subsp *thermophilus* and *Lactobacillus delbrueckii* subsp *bulgaricus*). Bacterial enzymes synthesized from these bacteria are thought to be responsible for the improved lactose digestion.<sup>80,81</sup> Researchers hypothesized that different strains and species of lactic acid bacteria would digest lactose more or less efficiently due to their varying activity of bacterial enzymes. When they tested different yogurts on seven lactase-deficient subjects, they found that all yogurts improved lactose digestion (despite their varying bacterial enzyme activity), but that different lactic acid bacteria strains resulted in different levels of improved lactose digestion. For example, *B. bifidus* milk gave the subjects only minimal improvement, while *L. bulgaricus* milk resulted in almost complete lactose digestion.<sup>79</sup>

## INFANT ALLERGIES

Alvarez first suggested in 1939 that poi be used as a substitute food for allergic people.<sup>82</sup> Poi is a potential probiotic found in the Pacific Islands made from the starchy corm of the Taro plant. During World War II, many military-associated people used poi as a substitute starch for people allergic to cereal or grain.<sup>83</sup> Later, Dr. Jerome Glaser, a pediatrician and allergist visiting Hawaii in 1961, observed that many infants in Hawaii were provided poi, especially infants with allergies or gastrointestinal problems.<sup>84</sup> He suggested that infants allergic to cereal could use poi as a substitute. Glaser reported that 19 rice-fed babies and 28 poi-fed babies had similar growth curves over a 2-month period. He also noted that only three of the 22 poi-fed babies (14%) had hematocrits that were 30 or less, compared with three out of 11 rice-fed infants (27%). Roth et al.<sup>85</sup> supported Glaser's findings when they concluded that poi was definitely well tolerated by babies, showing that poi may be regarded as a useful alternative when there is a family history of cereal allergy. Later, a study by Kalliomaki et al.<sup>86</sup> found that expectant mothers given a capsule of *Lactobacillus GG* for 2 weeks prior to delivery followed by their infants receiving the same capsule from birth to age 6 months resulted in half of the experimental infants developing eczema during that time period. The occurrence of eczema in infancy is a good indicator that a food allergy will develop later in childhood. These positive research results have prompted some researchers to suggest that probiotics might be a novel approach to treating food allergies.<sup>87</sup>

## FAILURE-TO-THRIVE

A few studies on the use of poi and failure-to-thrive were completed, but they date back to the mid 1960s. Glaser et al.<sup>84</sup> reported that 12 preterm infants consumed poi and thrived as well as other preterm infants (of comparable weight and size). A case study of a failure-to-thrive premature infant weighing 1500 grams noted that the infant was on various formulas but gained only 100 grams in 54 days. The infant's test results were all negative, including gastrointestinal x-rays, sweat electrolytes, carbohydrate utilization tests, and blood chemistry tests. She responded positively when provided poi and was able to maintain a healthy weight (2250 to 2500 grams). The authors therefore reported that poi could be safely recommended as a food for any very young infant, but some doctors question this because it is not sterilized.<sup>84</sup> These studies also occurred more than 50 years ago, and recommendations for future probiotic research would include studies with failure-to-thrive infants.

## HYPERLIPIDIMIA

Human and animal studies have suggested that the use of dairy products fermented with probiotics (lactic acid bacteria and bifidobacteria) may reduce serum lipid levels.<sup>78,88–93</sup> Two studies with normal-lipidemic subjects reported that probiotic administration resulted in a reduction in serum triglycerides (19% and 27%, respectively), along with slight changes in serum total and LDL cholesterol.<sup>94</sup> The proposed mechanism by which probiotics may decrease serum cholesterol is suggested to be related to the fermentation of indigestible dietary carbohydrates. Products of bacterial fermentation, specifically short-chain fatty acids, may inhibit cholesterol synthesis in the liver and/or cause the mobilization of plasma cholesterol to the liver.<sup>94</sup> Some gastrointestinal bacteria may also prevent cholesterol absorption by deconjugating bile salts that then affect cholesterol metabolism. Taranto et al.<sup>95</sup> reported that administration of *Lactobacillus reuteri* was effective in preventing hypercholesterolemia in mice. In addition, he observed a decrease in total cholesterol (22%) and triglycerides (33%), as well as a 17% increase in the ratio of HDL to LDL. In a study conducted by Usman and Hosono,<sup>96</sup> *Lactobacillus gasseri* was shown to lower serum lipids in hypercholesterolemic rats receiving nonfermented milk produced from *L. gasseri*. Total cholesterol and LDL levels by 42 and 64%, respectively.

## HEPATIC DISEASES

Mechanisms by which probiotics may treat hepatic encephalopathy have been suggested to include the following: (1) decreased portal blood ammonia by reduced bacterial urease activity, decreased pH due to less ammonia absorption, less intestinal permeability and improved gut epithelium; (2) decreased inflammation and oxidative stress due to reduced ammonia and toxins; and (3) reduced uptake of other toxins.<sup>97</sup> Solga<sup>97</sup> reported that the probiotic combination of VSL#3 reduced stool urease activity and pH, altered the production of short-chain fatty acids, and decreased inflammation in colonic cells.

Probiotics may also be effective in treating nonalcoholic fatty liver disease (NAFLD). Li et al.<sup>98</sup> studied the effects of VSL#3 on 48 ob/ob mice with NAFLD. The results were an improved liver histology, decreased total fatty acid content of the liver, and reduced serum ALT levels.

## HELICOBACTER PYLORI INFECTIONS

Studies suggest that probiotics, especially lactic acid bacteria, could be effective in the treatment and prevention of *Helicobacter pylori*, the bacteria cited as a causative agent of ulcers. In vitro studies have suggested that lactic acid bacteria may inhibit or kill *H. pylori* by acting as a bactericide.<sup>99,100</sup> Bifidobacteria and *B. subtilis* may inhibit the growth or attachment of *H. pylori*.<sup>101</sup> Cruchet et al.<sup>99</sup> conducted a double-blind, randomized, controlled

clinical trial to investigate the effects of *H. pylori* colonization in children given *Lactobacillus johnsonii* La 1, which interfered with *H. pylori* colonization by restricting the size of the pathogen's population and delaying colonization. Sakamoto et al.<sup>102</sup> also found that *H. pylori* could be eradicated in gnotobiotic murine models administered *Lactobacillus gasseri* OLL 2716(LG21). Possible mechanisms by which *L. salivarius* eradicates *H. pylori* include the ability of the former to bind to gastric epithelial cells, to produce a high quantity of lactic acid, and to proliferate rapidly.<sup>31</sup> Bhatia et al.<sup>103</sup> suggested the mechanism by which *L. acidophilus* may inhibit *H. pylori* is through the production of lactic acid.

## GENITOURINARY TRACT INFECTIONS

It has been suggested that some probiotics may be of benefit in the treatment and prevention of genitourinary tract infections such as vaginitis, urinary tract infections, and bacterial vaginosis.<sup>104–108</sup> According to Reid and Burton<sup>105</sup>, the probiotic *Lactobacillus* has the potential to prevent infections of the urogenital and intestinal tracts. Instillation of *Lactobacillus* GR-1 and B-54 or RC-14 directly into the vagina and oral ingestion of the probiotic have been shown to reduce the risk of urinary tract infections by creating a healthier environment within the vaginal flora. Specifically, lactic acid bacteria may protect and treat genitourinary infections because they are easily cultivated, non-pathogenic, population stable, and can adhere to vaginal epithelial cells,<sup>109</sup> forming a protective barrier to prevent colonization of pathogenic bacteria.<sup>110</sup>

## OTHER POSSIBLE USES OF PROBIOTICS

Other health conditions that may benefit from probiotic consumption include hypertension,<sup>111–115</sup> illness-related weight-loss,<sup>116,117</sup> and alcohol-induced liver damage.<sup>20,118</sup> Takano<sup>111</sup> suggested that probiotics can be used to treat hypertension when he reported that bioactive peptides produced from the proteolytic action of probiotic bacteria on casein—the protein found in milk—during milk fermentation may be able to lower the blood pressure of hypertensive individuals. Evidence to support his findings can be found in animal studies by Nakamura et al.<sup>112,113</sup> and one human study by Hata et al.,<sup>114</sup> which found that the dairy-based fermentation of milk by *Saccharomyces cerevisiae* and *Lactobacillus helveticus* resulted in the formation of two tripeptides that may have a role in lowering blood pressure. These two tripeptides, valine-proline-proline and isoleucine-proline-proline, act as angiotensin-I-converting enzyme inhibitors (ACE inhibitors) to lower blood pressure. Kalliomaki et al.<sup>86</sup> reported that perinatal administration of probiotics resulted in less atopic eczema in at-risk infants. These potential other benefits of probiotics remain inconclusive and controversial.

## CONCLUSION

Probiotics may play a beneficial role in several medical conditions, including diarrhea, gastroenteritis, irritable bowel syndrome, and inflammatory bowel disease (Crohn's disease and ulcerative colitis), cancer, depressed immune function, inadequate lactase digestion, infant allergies, failure-to-thrive, hyperlipidemia, hepatic diseases, *Helicobacter pylori* infections, genitourinary tract infections, and others, all of which are suggested by certain research studies to improve with the use of probiotics. Probiotics should be further investigated for their possible benefits to patients affected by these and possibly other medical conditions. At the same time, the potential for negative side effects from probiotics should also be researched. The correct combination and concentration of gastrointestinal microflora is determined by nature and numerous interdependent variables. Changing one factor such as concentration and trying to “optimize” nature's delicately balanced gastrointestinal environment may very well be altering a condition that nature never intended to alter. The short- and long-term effects of this change may be difficult to evaluate given the multifactorial nature of the gastrointestinal environment.



### Acknowledgements

This research was supported by a Research Centers in Minority Institutions award, P20 RR11091, from the National Center for Research Resources, National Institutes of Health. Its contents are solely the responsibility of the authors and do not necessarily represent the official views of the NCCR/NIH. A grant from the USDA-CSREES (Cooperative State Research, Education, and Extension Service) also supported this research.

### References

1. Marteau PR, de Vrese M, Cellier CJ, Schrezenmeir J. Protection from gastrointestinal diseases with the use of probiotics. *Am J Clin Nutr* 2001;73(suppl 2):430S–436S. [PubMed: 11157353]
2. Fuller R. Probiotics in human medicine. *Gut* 1991;32:439–442. [PubMed: 1902810]
3. Gismondo MR, Drago L, Lombardi A. Review of probiotics available to modify gastrointestinal flora. *Int J Antimicrob Agents* 1999;12:287–292. [PubMed: 10493604]
4. Salminen S, Isolauri E, Onnela T. Gut microflora in normal and disordered states. *Chemotherapy* 1995;41(suppl 1):5–15. [PubMed: 7671648]
5. Salminen S. Lactic acid bacteria in health and disease. *Lactic Acid Bacteria: Microbiology and Functional Aspects*. 2nd ed. New York: Marcel Dekker; 1998:211–254.
6. Schaafsma G. Application of lactic acid bacteria in novel foods from a nutritional perspective. *Les Bacteries Lactiques: Quelles Souches, Pour Quels Marches? Actes du Colloque LACTIC 94, Caen, 7–9 septembre 1994, pp.85–89*. Presses Universitaires de Caen; 1995:85–93.
7. Bengmark S. Colonic food: pre- and probiotics. *Am J Gastroenterol* 2000;95(suppl 2):S5–S7. [PubMed: 10634219]
8. de Roos NM, Katan MB. Effects of probiotic bacteria on diarrhea, lipid metabolism, and carcinogenesis: a review of papers published between 1988 and 1998. *Am J Clin Nutr* 2000;71:405–411. [PubMed: 10648252]
9. Batt RM, Rutgers HC, Sancak AA. Enteric bacteria: friend or foe? *J Small Anim Pract* 1996;37:261–267. [PubMed: 8805096]
10. Spanhaak S, Havenaar R, Schaafsma G. The effect of consumption of milk fermented by *Lactobacillus casei* strain Shirota on the intestinal microflora and immune parameters in humans. *Eur J Clin Nutr* 1998;52:899–907. [PubMed: 9881885]
11. Benno Y, Mitsuoka T. Impact of *Bifidobacterium longum* on human fecal microflora. *Microbiol Immunol* 1992;36:683–694. [PubMed: 1406371]
12. Ling WH, Korpela R, Mykkanen H, Salminen S, Hanninen O. *Lactobacillus* strain GG supplementation decreases colonic hydrolytic and reductive enzyme activities in healthy female adults. *J Nutr* 1994;124:18–23. [PubMed: 8283290]
13. Pahwa A, Mathur BN. Assessment of a bifidus containing infant formula. Part II. Implantation of *Bifidobacterium bifidum*. *Indian J Dairy Sci* 1987;40:364–367.
14. Langhendries JP, Detry J, Van Hees J, et al. Effect of a fermented infant formula containing viable bifidobacteria on the fecal flora composition and pH of healthy full-term infants. *J Pediatr Gastroenterol Nutr* 1995;21:177–181. [PubMed: 7472904]
15. Marteau P, Minekus M, Havenaar R, Huis in't Veld JH. Survival of lactic acid bacteria in a dynamic model of the stomach and small intestine: validation and the effects of bile. *J Dairy Sci* 1997;80:1031–1037. [PubMed: 9201571]
16. Pochart P, Marteau P, Bouhnik Y, Goderel I, Bourlioux P, Rambaud JC. Survival of bifidobacteria ingested via fermented milk during their passage through the human small intestine: an in vivo study using intestinal perfusion. *Am J Clin Nutr* 1992;55:78–80. [PubMed: 1728822]
17. Heller KJ. Probiotic bacteria in fermented foods: product characteristics and starter organisms. *Am J Clin Nutr* 2001;73(suppl 2):374S–379S. [PubMed: 11157344]
18. Holzapfel WH, Haberer P, Geisen R, Bjorkroth J, Schillinger U. Taxonomy and important features of probiotic microorganisms in food and nutrition. *Am J Clin Nutr* 2001;73(suppl 2):365S–373S. [PubMed: 11157343]
19. Elmer GW, Surawicz CM, McFarland LV. Bio-therapeutic agents: a neglected modality for the treatment and prevention of selected intestinal and vaginal infections. *JAMA* 1996;275:870–876. [PubMed: 8596226]

20. Goldin BR. Health benefits of probiotics. *Br J Nutr* 1998;80:S203–S207. [PubMed: 9924285]
21. Gorbach SL. Probiotics and gastrointestinal health. *Am J Gastroenterol* 2000;95(suppl 1):S2–S4. [PubMed: 10634218]
22. Black FT, Anderson PL, Orskov J, Orskov F, Gaarslev K, Laulund S. Prophylactic efficacy of lactobacilli on traveler's diarrhea. *Travel Med* 1989;7:333–335.
23. Gionchetti P, Rizzello F, Venturi A, et al. Oral bacteriotherapy as maintenance treatment in patients with chronic pouchitis: a double-blind, placebo-controlled trial. *Gastroenterology* 2000;119:305–309. [PubMed: 10930365]
24. Hilton E, Kolakowski P, Singer C, Smith M. Efficacy of *Lactobacillus* GG as a diarrheal preventive in travelers. *J Travel Med* 1997;4:41–43. [PubMed: 9815476]
25. Rolfe RD. The role of probiotic cultures in the control of gastrointestinal health. *J Nutr* 2000;130 (suppl 2S):396S–402S. [PubMed: 10721914]
26. Guandalini S, Pensabene L, Zikri MA, et al. Lactobacillus GG administered in oral rehydration solution to children with acute diarrhea: a multicenter European trial. *J Pediatr Gastroenterol Nutr* 2000;30:54–60. [PubMed: 10630440]
27. Van Niel CW, Feudtner C, Garrison MM, Christakis DA. *Lactobacillus* therapy for acute infectious diarrhea in children: a meta-analysis. *Pediatrics* 2002;109:678–684. [PubMed: 11927715]
28. Clements ML, Levine MM, Black RE, et al. Lactobacillus prophylaxis for diarrhea due to enterotoxigenic *Escherichia coli*. *Antimicrob Agents Chemother* 1981;20:104–108. [PubMed: 6792978]
29. Camarri E, Belvisi A, Guidoni G, Marini G, Frigerio G. A double-blind comparison of two different treatments for acute enteritis in adults. *Chemotherapy* 1981;27:466–470. [PubMed: 7028412]
30. Midolo PD, Lambert JR, Hull R, Luo F, Grayson ML. In vitro inhibition of *Helicobacter pylori* NCTC 11637 by organic acids and lactic acid bacteria. *J Appl Bacteriol* 1995;79:475–479. [PubMed: 7592140]
31. Aiba Y, Suzuki N, Kabir AM, Takagi A, Koga Y. Lactic acid-mediated suppression of *Helicobacter pylori* by the oral administration of *Lactobacillus salivarius* as a probiotic in a gnotobiotic murine model. *Am J Gastroenterol* 1998;93:2097–2101. [PubMed: 9820379]
32. Kabir AM, Aiba Y, Takagi A, et al. Prevention of *Helicobacter pylori* infection by lactobacilli in a gnotobiotic murine model. *Gut* 1997;41:49–55. [PubMed: 9274471]
33. Gibson GR, Saavedra JM, MsFarlaneS, McFarlane GT. Probiotics and intestinal infections. In: Fuller R., ed. *Probiotics 2: Applications and Practical Aspects*, 2nd ed. London: Chapman & Hall; 1997:10–38.
34. Marteau P, Pochart P, Bouhnik Y, Rambaud JC. The fate and effects of transiting, nonpathogenic microorganisms in the human intestine. *World Rev Nutr Diet* 1993;74:1–21. [PubMed: 8212727]
35. Nobaek S, Johansson ML, Molin G, Ahme S, Jeppsson B. Alteration of intestinal microflora is associated with reduction in abdominal bloating and pain in patients with irritable bowel syndrome. *Am J Gastroenterol* 2000;95:1231–1238. [PubMed: 10811333]
36. Niedzielin K, Kordecki H, Birkenfeld B. A controlled, double-blind, randomized study on the efficacy of *Lactobacillus plantarum* 299V in patients with irritable bowel syndrome. *Eur J Gastroenterol Hepatol* 2001;13:1143–1147. [PubMed: 11711768]
37. Kim HJ, Camilleri M, McKinzie S, et al. A randomized controlled trial of a probiotic, VSL#3, on gut transit and symptoms in diarrhoea-predominant irritable bowel syndrome. *Aliment Pharmacol Ther* 2003;17:895–904. [PubMed: 12656692]
38. Brigidi P, Vitali B, Swennen E, Bazzocchi G, Matteuzzi D. Effects of probiotic administration upon the composition and enzymatic activity of human fecal microbiota in patients with irritable bowel syndrome or functional diarrhea. *Res Microbiol* 2001;152:735–741. [PubMed: 11686387]
39. Gionchetti P, Rizzello F, Venturi A, Campieri M. Probiotics in infective diarrhoea and inflammatory bowel diseases. *J Gastroenterol Hepatol* 2000;15:489–493. [PubMed: 10847433]
40. Fabia R, Ar'Rajab A, Johansson ML, et al. Impairment of bacterial flora in human ulcerative colitis and experimental colitis in the rat. *Digestion* 1993;54:248–255. [PubMed: 8243838]
41. Sartor RB. Current concepts of the etiology and pathogenesis of ulcerative colitis and Crohn's disease. *Gastroenterol Clin North Am* 1995;24:475–507. [PubMed: 8809232]

42. Schultz M, Sartor RB. Probiotics and inflammatory bowel diseases. *Am J Gastroenterol* 2000;95 (suppl 1):S19–S21. [PubMed: 10634224]
43. Malin M, Suomalainen H, Saxelin M, Isolauri E. Promotion of IgA immune response in patients with Crohn's disease by oral bacteriotherapy with *Lactobacillus* GG. *Ann Nutr Metab* 1996;40:137–145. [PubMed: 8862696]
44. Plein K, Hotz J. Therapeutic effects of *Saccharomyces boulardii* on mild residual symptoms in a stable phase of Crohn's disease with special respect to chronic diarrhea—a pilot study. *Z Gastroenterol* 1993;31:129–134. [PubMed: 8465554]
45. Drisko JA, Giles CK, Bischoff BJ. Probiotics in health maintenance and disease prevention. *Altern Med Rev* 2003;8:143–155. [PubMed: 12777160]
46. Malchow HA. Crohn's disease and *Escherichia coli*. A new approach in therapy to maintain remission of colonic Crohn's disease? *J Clin Gastroenterol* 1997;25:653–658. [PubMed: 9451682]
47. Venturi A, Gionchetti P, Rizzello F, et al. Impact on the composition of the faecal flora by a new probiotic preparation: preliminary data on maintenance treatment of patients with ulcerative colitis. *Aliment Pharmacol Ther* 1999;13:1103–1108. [PubMed: 10468688]
48. Rafter JJ. The role of lactic acid bacteria in colon cancer prevention. *Scand J Gastroenterol* 1995;30:497–502. [PubMed: 7569753]
49. Goldin BR, Swenson L, Dwyer J, Sexton M, Gorbach SL. Effect of diet and *Lactobacillus acidophilus* supplements on human fecal bacterial enzymes. *J Natl Cancer Inst* 1980;64:255–261. [PubMed: 6766508]
50. Goldin BR. Intestinal microflora: metabolism of drugs and carcinogens. *Ann Med* 1990;22:43–48. [PubMed: 1970483]
51. Marteau P, Pochart P, Flourie B, et al. Effect of chronic ingestion of a fermented dairy product containing *Lactobacillus acidophilus* and *Bifidobacterium bifidum* on metabolic activities of the colonic flora in humans. *Am J Clin Nutr* 1990;52:685–688. [PubMed: 2119557]
52. Pedrosa MC, Golner BB, Goldin BR, Barakat S, Dallal GE, Russell RM. Survival of yogurt-containing organisms and *Lactobacillus gasseri* (ADH) and their effect on bacterial enzyme activity in the gastrointestinal tract of healthy and hypochlorhydric elderly subjects. *Am J Clin Nutr* 1995;61:353–359. [PubMed: 7840074]
53. Hayatsu H, Hayatsu T. Suppressing effect of *Lactobacillus casei* administration on the urinary mutagenicity arising from ingestion of fried ground beef in the human. *Cancer Lett* 1993;73:173–179. [PubMed: 8221630]
54. Lee YK, Salminen S. The coming age of probiotics. *Trends Food Sci Technol* 1995;6:241–245.
55. Lidbeck A, Nord CE, Gustafsson JA, Rafter J. Lactobacilli, anticarcinogenic activities and human intestinal microflora. *Eur J Cancer Prev* 1992;1:341–353. [PubMed: 1463986]
56. Burns AJ, Rowland IR. Anti-carcinogenicity of probiotics and prebiotics. *Curr Issues Intest Microbiol* 2000;1:13–24. [PubMed: 11709850]
57. Aso Y, Akazan H. Prophylactic effect of a *Lactobacillus casei* preparation on the recurrence of superficial bladder cancer. BLP Study Group. *Urol Int* 1992;49:125–129. [PubMed: 1466089]
58. Aso Y, Akaza H, Kotake T, Tsukamoto T, Imai K, Naito S. Preventive effect of a *Lactobacillus casei* preparation on the recurrence of superficial bladder cancer in a double-blind trial. The BLP Study Group. *Eur Urol* 1995;27:104–109. [PubMed: 7744150]
59. Orrhage K, Sillerstrom E, Gustafsson JA, Nord CE, Rafter J. Binding of mutagenic heterocyclic amines by intestinal and lactic acid bacteria. *Mutat Res* 1994;311:239–248. [PubMed: 7526189]
60. Lidbeck A, Nord CE, Rafter J, Nord CE, Gustafsson JA. Effect of *Lactobacillus acidophilus* supplements on mutagen excretion in faeces and urine in humans. *Microbiol Ecol Health Dis* 1992;5:59–67.
61. Goldin BR, Gualtieri LJ, Moore RP. The effect of *Lactobacillus* GG on the initiation and promotion of DMH-induced intestinal tumors in the rat. *Nutr Cancer* 1996;25:197–204. [PubMed: 8710689]
62. Wollowski I, Rechkemmer G, Pool-Zobel BL. Protective role of probiotics and prebiotics in colon cancer. *Am J Clin Nutr* 2001;73(suppl 2):451S–455S. [PubMed: 11157356]
63. Hirayama K, Rafter J. The role of probiotic bacteria in cancer prevention. *Microbes Infect* 2000;2:681–686. [PubMed: 10884619]

64. Vanderhoof JA. Probiotics: future directions. *Am J Clin Nutr* 2001;73:1152S–1155S. [PubMed: 11393194]
65. Biffi A, Coradini D, Larsen R, Riva L, Di Fronzo G. Antiproliferative effect of fermented milk on the growth of a human breast cancer cell line. *Nutr Cancer* 1997;28:93–99. [PubMed: 9200156]
66. Goldin BR, Gorbach SL. The effect of milk and lactobacillus feeding on human intestinal bacterial enzyme activity. *Am J Clin Nutr* 1984;39:756–761. [PubMed: 6424430]
67. Modler GW, McKellar RC, Yaguchi M. Bifidobacteria and bifidogenic factors. *Can Inst Food Sci Technol J* 1990;23:29–41.
68. Biasco G, Paganelli GM, Brandi G, et al. Effect of *Lactobacillus acidophilus* and *Bifidobacterium bifidum* on rectal cell kinetics and fecal pH. *Ital J Gastroenterol* 1991;23:142. [PubMed: 1742509]
69. Erickson KL, Hubbard NE. Probiotic immuno-modulation in health and disease. *J Nutr* 2000;130 (suppl 2S):403S–409S. [PubMed: 10721915]
70. Perdigon G, Alvarez S, Pesce de Ruiz Holgado A. Immunoadjuvant activity of oral *Lactobacillus casei*: influence of dose on the secretory immune response and protective capacity in intestinal infections. *J Dairy Res* 1991;58:485–496. [PubMed: 1722492]
71. Perdigon G. Probiotics and the immune state, In: Fuller R. *Probiotics: The Scientific Basis*. London: Chapman & Hall; 1992:145.
72. De Simone C, Vesely R, Bianchi-Salvadori B, Jirillo E. The role of probiotics in modulation of the immune system in man and in animals. *Int J Immunother* 1993;9:23–28.
73. Kolars JC, Levitt MD, Aouji M, Savaiano SA. Yogurt: an autodigesting source of lactose. *N Engl J Med* 1984;310:1–3. [PubMed: 6417539]
74. Savaiano DA, AbouElAnouar A, Smith DE, Levitt MD. Lactose malabsorption from yogurt, pasteurized yogurt, sweet acidophilus milk, and cultured milk in lactase-deficient individuals. *Am J Clin Nutr* 1984;40:1219–1223. [PubMed: 6439026]
75. Martini MC, Bollweg GL, Levitt MD, Savaiano DA. Lactose digestion by yogurt beta-galactosidase: influence of pH and microbial cell integrity. *Am J Clin Nutr* 1987;45:432–436. [PubMed: 3101480]
76. Dewit O, Boudraa G, Touhami M, Desjeux JF. Breath hydrogen test and stools characteristics after ingestion of milk and yogurt in malnourished children with chronic diarrhoea and lactase deficiency. *J Trop Pediatr* 1987;33:177–180. [PubMed: 3669132]
77. Lerebours E, N'Djitoyap Ndam C, Lavoine A, Hellot MF, Antoine JM, Colin R. Yogurt and fermented-then-pasteurized milk: effects of short-term and long-term ingestion on lactose absorption and mucosal lactase activity in lactase-deficient subjects. *Am J Clin Nutr* 1989;49:823–827. [PubMed: 2497632]
78. Marteau P, Flourie B, Pochart P, et al. Direct measurement of the absorption of lactose in yogurt in lactase-deficient men and the study of the fate of bacterial lactase in yogurt in the intestine. In *Fermented Milks: Current Research*. Paris: Syndifrais and the International Association of Yogurt Manufacturers; 1989:179–182.
79. Martini MC, Lerebours EC, Lin WJ, et al. Strains and species of lactic acid bacteria in fermented milks (yogurts): effect on in vivo lactose digestion. *Am J Clin Nutr* 1991;54:1041–1046. [PubMed: 1957819]
80. Premi L, Sandine WE, Elliker PR. Lactose-hydrolyzing enzymes of *Lactobacillus* species. *Appl Microbiol* 1972;24:51–57. [PubMed: 5057373]
81. Rao MV, Dutta SM. Purification and properties of B-galactosidase from *Streptococcus thermophilus*. *J Food Sci* 1981;46:1419–1423.
82. Alvarez W. Problems of maintaining nutrition in the highly food-sensitive person. *Am J Digest Dis* 1939;5:801–803.
83. Derstine V, Rada E. Some dietetic factors influencing the market for poi in Hawaii. In: *Agricultural Economics*, bulletin 3. University of Hawaii Agricultural Experiment Station, Hawaii. 1952;3:1–43.
84. Glaser J, Lawrence RA, Harrison A, Ball MR. Poi—its use as a food for normal, allergic and potentially allergic children. *Ann Allergy* 1967;25:496–500. [PubMed: 6069584]
85. Roth A, Worth RM, Lichten IJ. Use of poi in the prevention of allergic disease in potentially allergic infants. *Ann Allergy* 1967;25:501–506. [PubMed: 6069585]

86. Kalliomaki M, Salminen S, Poussa T, Arvilommi H, Isolauri E. Probiotics and prevention of atopic disease: 4-year follow-up of a randomised placebo-controlled trial. *Lancet* 2003;361:1869–1871. [PubMed: 12788576]
87. Majamaa H, Isolauri E. Probiotics: a novel approach in the management of food allergy. *J Allergy Clin Immunol* 1997;99:179–185. [PubMed: 9042042]
88. Khedkar CD, Garge RD, Mantri JM, Kulkarni SA, Khedkar GD. Effect of feeding acidophilus milk on serum cholesterol in human volunteers of 50 – 60 years. *J Dairy Foods Home Sci* 1993;12:33–38.
89. Lin SY, Ayres JW, Winkler W Jr, Sandine WE. Lactobacillus effects on cholesterol: in vitro and in vivo results. *J Dairy Sci* 1989;72:2885–2899. [PubMed: 2516523]
90. Schaafsma G, Meuling WJ, van Dokkum W, Bouley C. Effects of a milk product, fermented by *Lactobacillus acidophilus* and with fructo-oligosaccharides added, on blood lipids in male volunteers. *Eur J Clin Nutr* 1998;52:436–440. [PubMed: 9683397]
91. Agerbaek M, Gerdes LU, Richelsen B. Hypocholesterolaemic effect of a new fermented milk product in healthy middle-aged men. *Eur J Clin Nutr* 1995;49:346–352. [PubMed: 7664720]
92. Richelsen B, Kristensen K, Pedersen SB. Long-term (6 months) effect of a new fermented milk product on the level of plasma lipoproteins—a placebo-controlled and double blind study. *Eur J Clin Nutr* 1996;50:811–815. [PubMed: 8968702]
93. Sanders ME. Considerations for use of probiotic bacteria to modulate human health. *J Nutr* 2000;130 (suppl 2S):384S–390S. [PubMed: 10721912]
94. Pereira DI, Gibson GR. Effects of consumption of probiotics and prebiotics on serum lipid levels in humans. *Crit Rev Biochem Mol Biol* 2002;37:259–281. [PubMed: 12236466]
95. Taranto MP, Medici M, Perdigon G, et al. Effect of *Lactobacillus reuteri* on the prevention of hypercholesterolemia in mice. *J Dairy Sci* 2000;83:401–403. [PubMed: 10750094]
96. Usman, Hosono A. Effect of administration of *Lactobacillus gasseri* on serum lipids and fecal steroids in hypercholesterolemic rats. *J Dairy Sci* 2000;83:1705–1711. [PubMed: 10984145]
97. Solga SF. Probiotics can treat hepatic encephalopathy. *Med Hypotheses* 2003;61:307–313. [PubMed: 12888324]
98. Li Z, Yang S, Lin H, et al. Probiotics and antibodies to TNF inhibit inflammatory activity and improve nonalcoholic fatty liver disease. *Hepatology* 2003;37:343–350. [PubMed: 12540784]
99. Cruchet S, Obregon MC, Salazar G, Diaz E, Gotteland M. Effect of the ingestion of a dietary product containing *Lactobacillus johnsonii* La 1 on *Helicobacter pylori* colonization in children. *Nutrition* 2003;19:716–721. [PubMed: 12921879]
100. Hamilton-Miller JM. The role of probiotics in the treatment and prevention of *Helicobacter pylori* infection. *Int J Antimicrob Agents* 2003;22:360–366. [PubMed: 14522098]
101. Sullivan A, Nord CE. The place of probiotics in human intestinal infections. *Int J Antimicrob Agents* 2002;20:313–319. [PubMed: 12431865]
102. Sakamoto I, Igarashi M, Kimura K, Takagi A, Miwa T, Koga Y. Suppressive effect of *Lactobacillus gasseri* OLL 2716 (LG21) on *Helicobacter pylori* infection in humans. *J Antimicrob Chemother* 2001;47:709–710. [PubMed: 11328791]
103. Bhatia SJ, Kochar N, Abraham P, et al. *Lactobacillus acidophilus* inhibits growth of *Campylobacter pylori* in vitro. *J Clin Microbiol* 1989;27:2328–2330. [PubMed: 2511224]
104. Shalev E, Battino S, Weiner E, Colodner R, Keness Y. Ingestion of yogurt containing *Lactobacillus acidophilus* compared with pasteurized yogurt as prophylaxis for recurrent Candidal vaginitis and bacterial vaginosis. *Arch Fam Med* 1996;5:593–596. [PubMed: 8930233]
105. Reid G, Burton J. Use of *Lactobacillus* to prevent infection by pathogenic bacteria. *Microbes Infect* 2002;4:319–324. [PubMed: 11909742]
106. Hilton E, Isenberg HD, Alperstein P, France K, Borenstein MT. Ingestion of yogurt containing *Lactobacillus acidophilus* as prophylaxis for Candidal vaginitis. *Ann Intern Med* 1992;116:353–357. [PubMed: 1736766]
107. Hilton E, Rindos P, Isenberg HD. *Lactobacillus* GG vaginal suppositories and vaginitis. *J Clin Microbiol* 1995;33:1433. [PubMed: 7615776]

108. Hallen A, Jarstrand C, Pahlson C. Treatment of bacterial vaginosis with lactobacilli. *Sex Transm Dis* 1992;19:146–148. [PubMed: 1523530]
109. Barbes C, Boris S. Potential role of lactobacilli as prophylactic agents against genital pathogens. *AIDS Patient Care STDS* 1999;13:747–751. [PubMed: 10743538]
110. Ocana V, Nader-Macias ME. Adhesion of *Lactobacillus* vaginal strains with probiotic properties to vaginal epithelial cells. *Biocell* 2001;25:265–273. [PubMed: 11813542]
111. Takano T. Milk derived peptides and hypertension reduction. *Int Dairy J* 1998;8:375–381.
112. Nakamura Y, Yamamoto N, Sakai K, Takano T. Antihypertensive effect of sour milk and peptides isolated from it that are inhibitors to angiotensin I-converting enzyme. *J Dairy Sci* 1995;78:1253–1257. [PubMed: 7673515]
113. Nakamura Y, Masuda O, Takano T. Decrease of tissue angiotensin I-converting enzyme activity upon feeding sour milk in spontaneously hypertensive rats. *Biosci Biotechnol Biochem* 1996;60:488–489. [PubMed: 8901109]
114. Hata Y, Yamamoto M, Ohni M, Nakajima K, Nakamura Y, Takano T. A placebo-controlled study of the effect of sour milk on blood pressure in hypertensive subjects. *Am J Clin Nutr* 1996;64:767–771. [PubMed: 8901799]
115. Sawada H, Furushiro M, Hirai K, Motoike M, Watanabe T, Yokokura T. Purification and characterization of an antihypertensive compound from *Lactobacillus casei*. *Agric Biol Chem* 1990;54:3211–3219. [PubMed: 1368639]
116. Morrissey PJ, Charrier K, Braddy S, Liggitt D, Watson JD. CD4+ T cells that express high levels of CD45RB induce wasting disease when transferred into congenic severe combined immunodeficient mice. Disease development is prevented by cotransfer of purified CD4+ T cells. *J Exp Med* 1993;178:237–244. [PubMed: 8100269]
117. Aranda R, Sydora BC, McAllister PL, et al. Analysis of intestinal lymphocytes in mouse colitis mediated by transfer of CD4+, CD45RBhigh T cells to SCID recipients. *J Immunol* 1997;158:3464–3473. [PubMed: 9120308]
118. Nanji AA, Khettry U, Sadrzadeh SM. *Lactobacillus* feeding reduces endotoxemia and severity of experimental alcoholic liver (disease). *Proc Soc Exp Biol Med* 1994;205:243–247. [PubMed: 8171045]

Table 1

Commonly Used Bacterial Strains for Probiotic Purposes

<i>Lactobacillus</i> Species	<i>Bifidobacterium</i> Species	Other Lactic Acid Bacteria	Non-Lactic Acid Bacteria
<i>L. acidophilus</i> <i>L. bulgaricus</i> <i>L. casei</i> <i>L. rhamnosus</i> GG <i>L. plantarum</i>	<i>B. adolescentis</i> <i>B. animalis</i> <i>B. bifidum</i> <i>B. breve</i> <i>B. infantis</i> <i>B. longum</i> <i>B. thermophilus</i>	<i>Enterococcus faecium</i> <i>Streptococcus thermophilus</i>	<i>Bacillus subtilis</i> <i>Escherichia coli</i> strain nissle <i>Saccharomyces boulardii</i> <i>Saccharomyces cerevisiae</i>

From reference 3 and 18

**Table 2**  
Possible Probiotic Benefits (adapted from Goldin<sup>20</sup>)

---

Intestinal Disorders
Diarrhea
Antibiotic-associated
Traveler's
Pathogen-induced or infectious
Gastroenteritis
Irritable bowel syndrome
Inflammatory bowel disease
Crohn's disease
Ulcerative colitis
Pouchitis
Lactase digestion
Other Medical Disorders
Cancer
<i>Helicobacter pylori</i> infections
Hepatic diseases
Hyperlipidemia
Genitourinary tract infections
Improved immune function
Food substitute in allergies
Nutritional Supplement for Weight Gain
Failure-to-thrive
Cancer cachexia
AIDS
Pancreatitis/cystic fibrosis
Inflammatory bowel disease

---



Table 3  
 Clinical Trials Showing Significant Therapeutic Effects of Probiotics in Prevention of Antibiotic-associated Diarrhea (from Marteau et al.<sup>1</sup>)

Study	Antibiotic	Probiotic	Study Size	Blind Study	Probiotic Group vs. Control Group
Gotz, 1979	Ampicillin	<i>Lactobacillus acidophilus</i> + <i>Lactobacillus bulgaricus</i>	98	Yes	8.3% vs. 21%
Clements, 1983	Neomycin	<i>L. acidophilus</i> + <i>L. bulgaricus</i>	39	No	20% vs. 42%
Witsell, 1995	Amoxicillin-clavulanate	<i>L. acidophilus</i> + <i>L. bulgaricus</i>	27	No	Positive*
Borgia, 1982	Antituberculous	<i>Enterococcus faecium</i> SF68	200	No	5% vs. 18%
Wunderlich, 1989	Miscellaneous	<i>E. faecium</i> SF68	45	Yes	8.7% vs. 27.2%
Colombel, 1987	Erythromycin	<i>Bifidobacterium longum</i>	10	Yes	Positive*
Sitonen, 1990	Erythromycin	<i>Lactobacillus rhamnosus</i> GG	16	No	Positive*
Young, 1997	Miscellaneous	<i>L. rhamnosus</i> GG	188	No	17% vs. 48%
Ornhage, 1994	Clindamycin	<i>B. longum</i> + <i>Lactobacillus</i>	10	Yes	Positive
Adam, 1977	$\beta$ -lactams or tetracyclins	<i>Saccharomyces boulardii</i>	388	Yes	4.5% vs. 17.5%
Surawicz, 1989	Miscellaneous	<i>S. boulardii</i>	180	Yes	9.5% vs. 21.8%
McFarland, 1995	$\beta$ -lactams	<i>S. boulardii</i>	193	Yes	7.2% vs. 14.6%

\* The authors reported a positive effect of the probiotic but did not provide the percentage of subjects with antibiotic-associated adverse effects in the two groups.

**Table 4**  
 Clinical Trials of Probiotics Used to Prevent Traveler's Diarrhea (from Marteau et al.<sup>1</sup>)

Study	Probiotic	Study Size	Probiotic Group vs. Control Group
Pozo-Olano, 1978	<i>Lactobacillus acidophilus</i> + <i>Lactobacillus bulgaricus</i>	50	35% vs. 29% (NS)
Kollaritsch, 1983	Lactobacilli	212	55% vs. 51% (NS)
Katelaris, 1995	<i>Lactobacillus fermentum</i> strain KLD	282	23.8% vs. 23.8% (NS)
Katelaris, 1995	<i>L. acidophilus</i>	282	25.7% vs. 23.8% (NS)
Black, 1989	<i>L. acidophilus</i> + <i>Streptococcus thermophilus</i> + <i>Bifidobacterium bifidum</i> + <i>L. bulgaricus</i>	81	43% vs. 71% ( $P = 0.02$ )
Kollaritsch von, 1993	<i>Saccharomyces boulardii</i>	1016	28.7% vs. 39.1% ( $P < 0.05$ )
Oksaneva 1990	<i>Lactobacillus rhamnosus</i> GG	756	41% vs. 46.5% ( $P = 0.065$ )
Hilton, 1997	<i>L. rhamnosus</i> GG	245	3.9%/day vs. 7.4%/day ( $P = 0.05$ )

Table 5  
 Clinical Trials on Infants Showing Significant Therapeutic Effects of Probiotics Used in Acute Gastroenteritis (from Marteau et al.<sup>1</sup>)

Study	Disorder	Probiotic	Study Size
Curative Treatment			
Isolaurl, 1991	Rotavirus-associated diarrhea	<i>Lactobacillus rhamnosus GG</i>	71
Kaila, 1992		<i>L. rhamnosus GG</i>	39
Majamaa, 1995		<i>L. rhamnosus GG</i>	49
Isolaurl, 1994		<i>L. rhamnosus GG</i>	42
Sugita 1994		<i>L. casei Shirota</i>	32
Raza, 1995		<i>L. rhamnosus GG</i>	32
Pant, 1996	Gastroenteritis	<i>L. rhamnosus GG</i>	26
Guarino, 1997		<i>L. rhamnosus GG</i>	100
Shornikova, 1997		<i>L. rhamnosus GG</i>	123
Guandalini, 2000		<i>L. rhamnosus GG</i>	287
Bellomo, 1980*		<i>L. rhamnosus GG</i>	104
Chapoy, 1966		<i>Enterococcus faecium SF68</i>	38
Shornikova 1997		<i>Saccharomyces boulardii</i>	66
Prevention			
Saavedra, 1994	Acute diarrhea or rotavirus	<i>Bifidobacterium bifidum</i> + <i>Streptococcus thermophilus</i>	55

\* G. Boudraa, unpublished results, 1996.